

## The Effect of Produce Conditions for Preparation of Potentiometric Carbon Paste Sensor for Determination of Midazolam in Pharmaceutical

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The construction and general performance of a novel modified carbon paste electrode for the determination of midazolam have been developed. Midazolam-tetraphenylborate (MZ-TPB) ion pairs have been prepared and used as electro active materials. The electrode shows a stable, potentiometric response for midazolam in the concentration range  $5 \times 10^{-3}$  -  $1 \times 10^{-5}$  M at 25 °C independent of pH in the range 3 – 5. The electrode passed near Nernstian cationic slope of  $57.3 \pm 0.2$  mV and lower detection limit of  $6 \times 10^{-7}$  M with a fast response time of 5-10s. Selectivity coefficients for midazolam relative to a number of interfering substances have been investigated. There is a negligible interference from the studied cations. These results have been obtained using the proposed electrodes by using a (HPLC) reference method showed that the ion-selective electrode technique is sensitive, reliable and can be used with very good accuracy and high % recovery without pretreatment procedures of the samples to minimize interfering matrix effects.

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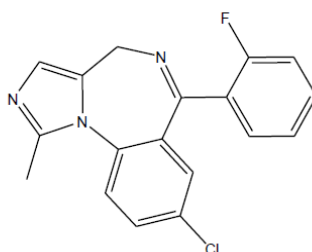
**Keywords:** Midazolam; Tetraphenylborate; carbon paste; potentiometric sensor

### 1. INTRODUCTION

Benzodiazepines are psychoactive therapeutic compounds possess sedative, hypnotic, anxiolytic, anticonvulsant, muscle relaxant, and amnesic actions [1, 2], which are useful in a variety of indications such as alcohol dependence, seizures, anxiety, panic, agitation and insomnia. They slow down the activity of the central nervous system. Benzodiazepines are classified as short, intermediate

or long-acting. Short and intermediate-acting benzodiazepines are preferred for the treatment of insomnia. Longer-acting benzodiazepines are recommended for the treatment of anxiety.

Midazolam (MDZ, 8-chloro-6-(2-fluorophenyl)-1-methyl-4H-imidazo-[1, 5-a] [1,4]-benzodiazepine)(fig.1) [3], is a benzodiazepine with a rapid onset and short duration of action [4,5]. MDZ commonly used in intravenous anesthesia induction, short-term sedation and oral hypnotic medication [6]. MDZ is basic and very stable in water solution [7]. The nitrogen in the 2-position provides sufficient basicity ( $pK_a = 6.15$ ). In strong acidic solutions the diazepine ring reversibly opens between the positions 4 and 5, producing a polar water-soluble primary amine derivative [8]. At physiological pH values, approximately 96% of MDZ is bound to plasma proteins [8].



**Figure 1.** Structural formula of midazolam.

Several analytical methods have been published for the determination of MDZ with or without its metabolites in different biological fluids. These techniques utilized high performance liquid chromatography (HPLC) with UV detection [9–15], HPLC–mass spectrometry (MS) [16–22], gas chromatography (GC)–MS [23, 24]. Although methods utilizing MS detectors are the most sensitive, and usually require small sample volume, these detectors are not always available in clinical settings and traditional research laboratories. Most of the HPLC–UV methods suffer from various limitations, including inadequate sensitivity [10–12, 14], long run times [12, 14], and use of expensive solid phase extraction cartridges [8, 9, 21]. This work is to introduce a very simple and inexpensive (potentiometric) method in determination of midazolam in a wide concentration range and in presence of variety of metal ions with minimum number of interferences. This method, potentiometric characterization, and analytical application of a novel midazolam-modified carbon paste electrode based on the use of midazolam-tetraphenylborate (MDZ-TPB) as electro active material and dioctylphthalate (DOP) as plasticizer.

## 2. EXPERIMENTAL

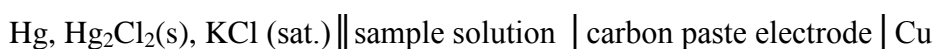
### 2.1. Materials

Pure midazolam, diazepam and oxzepam were purchased from Cambrex. Ampoule midazolam 5mg/ml was obtained from Tehran chemical pharmacy. All Reagents used were chemically pure grade and doubly distilled water was used throughout. Sodium tetraphenylborate (Na-TPB), graphite powder, acetic acid, sodium acetate, hydrochloric acid, mineral oil and methanol were obtained from Merck. O-

nitrophenyloctylether (o-NPOE) was obtained from Fluka. Bis(2-ethylhexyl)adipate (DOA), dioctylsebacate (DOS) and dioctyl phthalate (DOP) were obtained from Aldrich. All other solutions used in interference studies and electrode applicability were prepared from analytical grade salts (all from Aldrich or Merck).

## 2.2. Apparatus

All potentiometric measurements were made at  $25 \pm 1^\circ\text{C}$  unless otherwise stated using a Metrohm potentiometer (pH meter /Ion meter model of 781) and a combined Metrohm pH electrode (model 9101) was used for pH measurements. Saturated calomel electrode (SCE) was used as a reference electrode. The electrochemical system is represented as follows:



## 2.3. Standard solutions

### 2.3.1. Midazolam standard solutions

Stock solutions ( $1 \times 10^{-2}$  M) were prepared by dissolving the proper weight of 163mg midazolam drug into 20.0 ml  $1 \times 10^{-2}$  M HCl. A 2.0M HCl solution was added drop wise with continuous stirring till complete drug dissolution was achieved. The pH was adjusted to about 4 with dilute NaOH solution. The resulting solution was then made up to 50 ml in a measuring flask using the  $1 \times 10^{-2}$  M HCl. Working solutions of each drug in the concentration range ( $5 \times 10^{-3}$  to  $10^{-6}$  M) were prepared by serial accurate dilution with the  $1 \times 10^{-2}$  M HCl solution.

### 2.3.2. Tetraphenylborate solution (TPB)

A  $2 \times 10^{-2}$  M sodium tetraphenylborate solution was prepared by dissolving 342mg into 50ml de-ionized water. The resulting solution was standardized by potentiometric titration against  $2 \times 10^{-2}$  M silver nitrate solution using a silver/silver sulfide electrode in conjunction with a double-junction saturated calomel electrode.

### 2.3.3. Interfering ions solutions

A  $10^{-3}$  M standard solution each of cobalt nitrate, cadmium nitrate, calcium nitrate, nickel nitrate, aluminium nitrate, chromium nitrate, barium nitrate, iron sulfate, copper chloride, lead nitrate, zinc nitrate, diazepam, oxazepam, fructose, sucrose, galactoside and uric acid were prepared by dissolving the proper weights into 100 ml of de-ionized water. A 500 ppm starch solution was prepared by dissolving 50 mg into 100 ml de-ionized water.

#### 2.4. Preparation of midazolam-TPB ion pair complexes

A  $2 \times 10^{-2}$  M of midazolam drug solution was prepared by dissolving the proper weight into 25 ml aliquots of  $2 \times 10^{-2}$  M HCl. Equal volume of  $2 \times 10^{-2}$  M, tetraphenylborate (TPB) solution was added drop wise with continuous stirring. The resulting precipitate were left over night to settle down, filtered through G-4 sintered-glass crucibles, washed with distilled water till no chloride ion was detected in the filtrate and dried under vacuum for 48 h[

#### 2.5. Preparation of the midazolam-modified carbon paste electrode

The midazolam-modified carbon paste electrode (MDZCPE) was prepared by mixing weighed amounts of MDZ-/TPB as electro active material, graphite powder, plasticizer and methanol thoroughly until obtaining a uniformly wetted paste. Portions of the resulting Composite material were then packed in the end of a disposable polyethylene syringe (3 mm i.d., 1ml), the tip of which had been cut-off with a razor blade. Electrical contact to the carbon paste was made with a copper wire. Fresh surface was obtained by applying manual pressure to the carbon paste. The resulting fresh surface was polished on a filter paper until it had a shiny surface. The electrode surface was polished gently with smooth tissue of paper when the midazolam solution is changed from high concentration to a dilute solution. The newly prepared indicator electrode was conditioned by soaking in a  $1 \times 10^{-2}$  M aqueous midazolam solution for 1h and stored in the same solution when not in use.

#### 2.6. Procedure

The midazolam-modified carbon electrode was calibrated by immersion in conjunction with the reference electrode in a 50 ml beaker containing 9 ml of acetate buffer solution of pH 4. Then 1 ml aliquot of midazolam solution of concentration ranging from  $5 \times 10^{-3}$  to  $1 \times 10^{-6}$  M was added with continuous stirring and the potential was recorded after stabilization to  $\pm 0.2$  mV. A calibration graph was then constructed by plotting the recorded potentials as a function of  $-\log$  [midazolam]. The resulting graph was used for subsequent determination of unknown midazolam concentration.

#### 2.7. Determination of midazolam in midazolam ampoules

Midazolam was determined in midazolam injection solution (5mg/1 ml) by transferring the contents of one ampoule of midazolam into a 100 ml measuring flask, and making up to the mark with water. Fifty and hundred microliters aliquots of the solution were transferred to the measuring cell containing 9.0 ml of acetate buffer, and the e.m.f. of the electrode systems was measured. The concentration of midazolam was calculated from the previous calibration graph as in the procedure. Alternatively, the standard addition technique was used for the determination of midazolam by monitoring the potential of the drug solution before and after the addition of a known concentration of

the midazolam solution. The determination of midazolam content in midazolam ampoule has been carried out using HPLC methods [25] after appropriate dilution of the midazolam concentration.

### 2.8. Determination of midazolam in Biological Fluids

Different quantities of midazolam and 1mL serum were transferred to a 50mL measuring flask and completed to the mark with  $1 \times 10^{-4}$  M HCl to give solutions of pH values ranging from 4 and concentrations of  $5 \times 10^{-3}$  to  $1 \times 10^{-6}$  M midazolam. These solutions were subjected to the standard addition method for the potentiometric determination of midazolam.

### 2.9. Selectivity of the Sensor

Potentiometric selectivity factors of the electrode were evaluated by applying the matched potential method (MPM) [26]. According to this method, the activity of (CPM) was increased from  $a_A = 1.0 \times 10^{-4}$  M (reference solution) to  $a'_A = 2.0 \times 10^{-4}$  M, and the change in potential ( $\Delta E$ ) corresponding to this increase were measured. Next, a solution of an interfering ion of concentration  $a_B$  in the range  $1.0 \times 10^{-1}$  at  $1.0 \times 10^{-2}$  M is added to new  $1.0 \times 10^{-4}$  M (reference solution) until the same potential change ( $\Delta E$ ) was recorded. The selectivity factor,  $k_{A,B}^{MPM}$  for each interferent was calculated using the following equation:

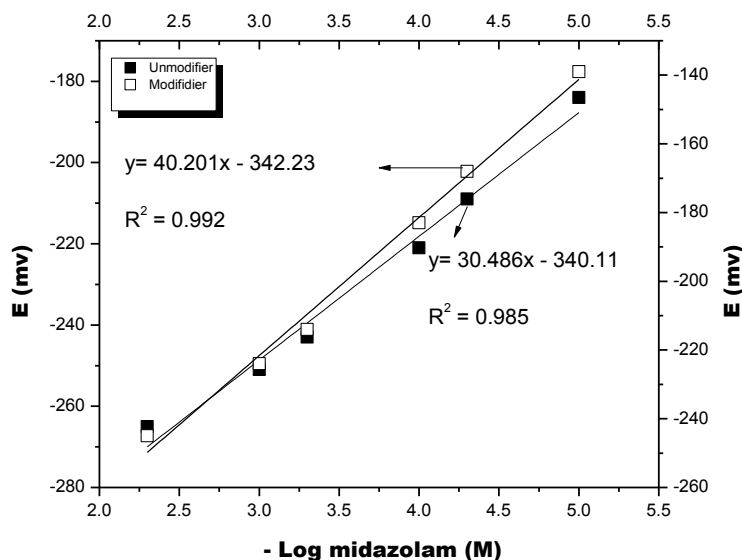
$$k_{A,B}^{MPM} = (a'_A - a_A) / a_B$$

## 3. RESULTS AND DISCUSSION

Tetraphenylborate was used as an ion-pairing agent for the preparation of an electro active ion association complex for midazolam. The elemental analysis of the sparingly soluble complex of MDZ-TPB showed that the composition of the complex is 1:1 (midazolam: tetraphenylborate). The dry powder of the formed ion-pair was used for the preparation of the new midazolam-modified carbon paste electrode [27-29]

### 3.1. Response characteristics of modified and unmodified carbon paste electrode

The unmodified electrode showed no significant response under the optimum conditions (Fig. 2). As can be seen, the electrode without ion-pair modifier and ion-pair modifier gave a working concentration range of  $5 \times 10^{-3}$  to  $1 \times 10^{-6}$  with a slope of 30.5 and 40.2 mV of the analyte.



**Figure 2.** Response of modified and unmodified carbon paste electrode under the optimum conditions. *Conditions:* buffer acetate with pH = 4, 5% modifier

3.2. Optimization of the amount of modifier in the electrode

It is known that the sensitivity and linearity for a given electrode depend significantly on the amount of the ion-pair in the membrane composition. Thus, the influence of the percent of MDZ-TPB in the carbon paste composition was investigated. Preliminary experiment showed that carbon paste electrode which does not contain the ion-pair modifier with slope 4.1 has no response towards the analyte. For this purpose, five electrodes were prepared that contain the ion-pair modifier in 5, 10, 12, 15, and 20%, while the other components have been kept unchanged, and the results are summarized in Table 1. The resulting slopes and correlation coefficients are 40.2 (0.992), 57.3 (0.998), 51.5 (0.993), 41.3 (.988) and 36.7 (0.992) mV per decade. These results show that for the electrode that contains 10.0% of the modifier, a Nernstian slope has been obtained. A nonlinearity in the electrode response has been observed with electrodes that contain less or higher ratios of the modifier. Since the electrode with 10 % of ion-pair has a good slope and wide range of linearity, this percentage was chosen as the optimum amount for the midazolam electrode.

**Table 1.** General characteristic of some different composition of MDZCPE

Composition of the modified chemical (%)	slope	Range of determination (M)	Lower LOD (M)	Correlation coefficient, r
5	40.1±0.2	5×10 <sup>-3</sup> - 1×10 <sup>-5</sup>	1.0×10 <sup>-6</sup>	0.992
10	57.3±0.2	5×10 <sup>-3</sup> - 1×10 <sup>-5</sup>	6.0×10 <sup>-7</sup>	0.998
12	51.5±0.3	5×10 <sup>-3</sup> - 1×10 <sup>-5</sup>	8.0×10 <sup>-7</sup>	0.993
15	41.3±0.3	5×10 <sup>-3</sup> - 1×10 <sup>-5</sup>	2.0×10 <sup>-6</sup>	0.988
20	36.7±0.4	5×10 <sup>-3</sup> - 1×10 <sup>-5</sup>	4.0×10 <sup>-6</sup>	0.992

*Conditions:* buffer acetate with pH = 4, plasticizer DOP 40%

3.3. Effect of plasticizer on the potential response

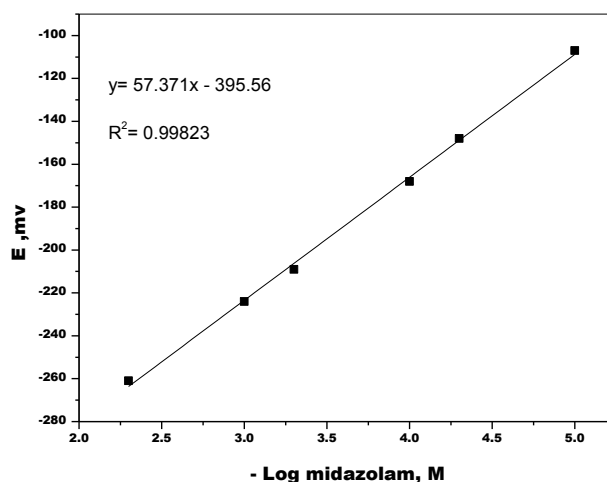
Next five electrodes were prepared that contain the ion-pair modifier 10%, Graphite powder 50% and different plasticizer (mineral oil, DOS, DOA, o-NPOE, and DOP) 40% the results are summarized in Table 2. The results show that for the electrode that contains DOP as plasticizer, a Nernstian slope has been obtained. Fig. 3 shows that the linear ranges of the electrode that contain ion-pair complex 10%, Graphite powder 50% and DOP plasticizer 40% is in the Range of  $5 \times 10^{-3}$  -  $1 \times 10^{-5}$  M. The limit of detection (LOD) is  $6.0 \times 10^{-7}$  M, respectively. The least-squares equation obtained from the calibration data is

$$E(mv) = -(57.3 \pm 0.2) \text{Log}[\text{midazolam}] - 395.57$$

**Table 2.** General characteristic of different plasticizer of MDZCPE

Composition of the plasticizer chemical (%)	slope	Range of determination (M)	Correlation coefficient, r
DOS	32.4±0.2	$5 \times 10^{-3}$ - $5 \times 10^{-5}$	0.973
DOP	57.3±0.2	$5 \times 10^{-3}$ - $1 \times 10^{-5}$	0.998
DOA	29.4±0.3	$5 \times 10^{-3}$ - $5 \times 10^{-5}$	0.988
o-NPOE	48.6±0.2	$5 \times 10^{-3}$ - $1 \times 10^{-5}$	0.945
mineral oil	40.8±0.3	$5 \times 10^{-3}$ - $5 \times 10^{-5}$	0.984

Conditions: buffer acetate with pH = 4, 10% modifier, 50% graphite powder, 40% plasticizer



**Figure 3.** Calibration graph

3.4. Repeatability or reproducibility

The repeatability of the method has been examined by measuring the potential response of different concentrations of midazolam over a wide time interval of 1 week. The repeatability of the measuring solution has been found to be within  $\pm 1$  mV over a week.

3.5. Effect of pH

The behavior of the electrode in relation to the variation of pH (2–7.5) at concentration of  $1 \times 10^{-4}$  M of midazolam was studied (Fig. 4). For measuring the pH, adjustments were made with the concentrated hydrochloric acid or sodium hydroxide solutions. It can be seen from Fig.4 that the variation in potential due to pH change is considered acceptable in the pH range 3 – 5. However, there is an observed drift at pH values lower than 3 which may be due to  $H^+$  interference. On the other hand, the potential decreases gradually at pH values higher than 5. This is possibly attributed to the midazolam the base form with lower solubility causing the electrodes potentials to decrease. The potentiometric curves of midazolam in buffer solutions with different pH values are illustrated in Fig. 5. According to Fig. 5 pH 4 offered a better slope and a wider linear range than in the other pH values.

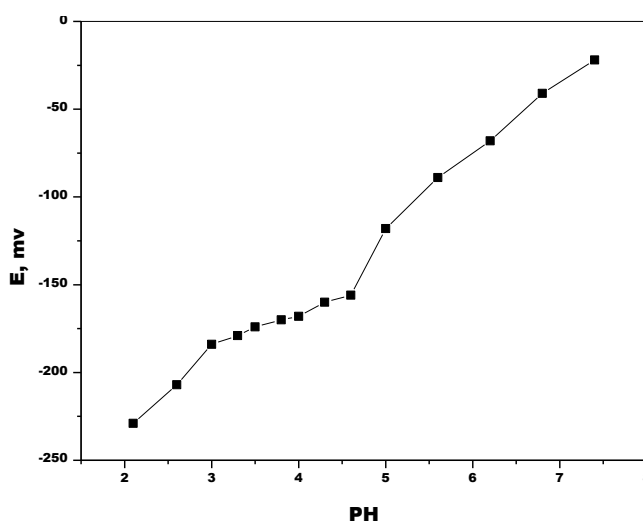


Figure 4. The pH effect on potential response of the midazolam electrode (conditions: 10 % ion-pair modifier, midazolam concentration was  $1.0 \times 10^{-4}$  M).

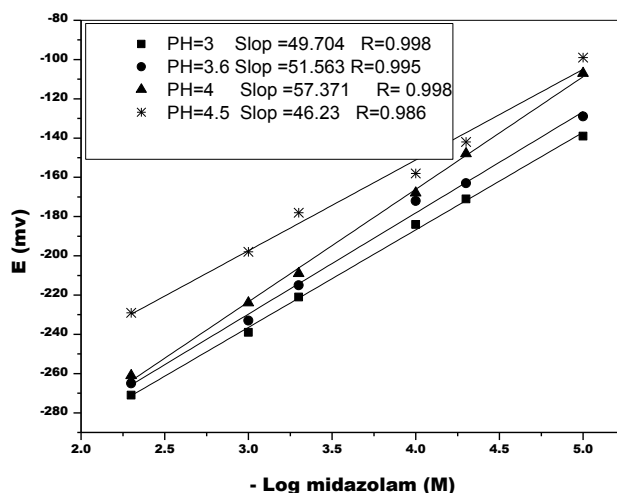


Figure 5. potentiometric curves of the proposed electrode at various pH buffers (conditions: 10% ion-pair modifier, midazolam concentration was  $1.0 \times 10^{-4}$  M)



3.6. Effect of response time

The dynamic response time of the modified electrode was measured according to IUPAC recommendation [30]. The response time of the electrode is defined as the time required for the electrode to reach a stable potential within  $\pm 1$  mV of the final equilibrium value after successive immersions of the sensor of a series of midazolam solutions, each having difference in concentration, was studied, The static response time of the midazolam carbon paste electrode was 3-8 s over the concentration range ( $5 \times 10^{-3}$  to  $1 \times 10^{-5}$  M) is shown in Figure 6, at lower concentrations, The response time was delayed and reached to 10 s and no change is observed up to 3 minutes. The repeatability of the potential reading of the electrode was examined by subsequent measurements (high to- low cycles) in  $1 \times 10^{-4}$  M midazolam solution immediately after measuring the first set of solutions at  $5 \times 10^{-5}$  M midazolam is show in figure 7.

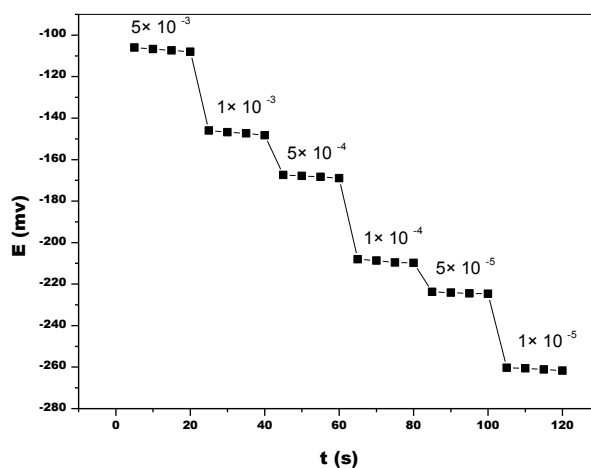


Figure 6. Dynamic response characteristics of midazolam carbon paste electrode for different concentration midazolam.

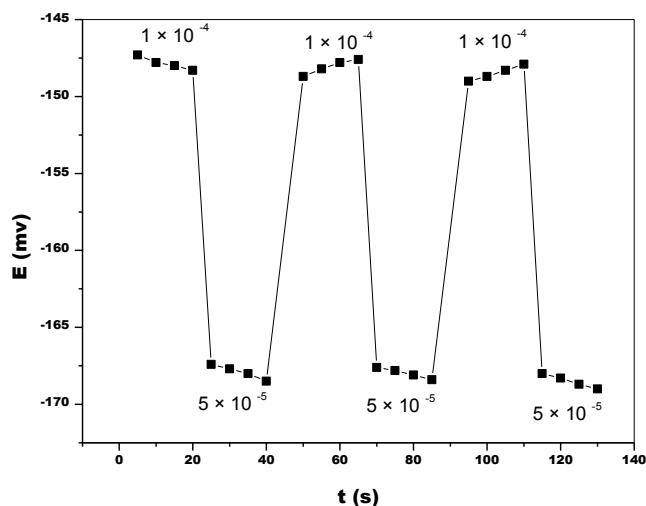


Figure 7. Dynamic response characteristics of midazolam carbon paste electrode for several high- to-low cycles.

### 3.7. Potentiometric Selectivity

The potentiometric selectivity coefficient of an electrode, as one of the most important characteristics, is defined by its relative response for the primary ion over the other ions present in the solution [31]. The selectivity coefficients of the modified carbon paste electrode towards many inorganic cations, drugs, carbohydrates were evaluated by the matched potential method (MPM) [25]. This method measures selectivity coefficients of ionic and nonionic species; it has an advantage of removing limitations imposed by Nikolsky –Eisenman equation while calculating selectivity coefficients by other methods. These limitations include non- Nernstian behavior of interfering ions and problems of inequality of charges of primary and interfering ions [32]. The values of the selectivity coefficients are listed in Table 3; they reflect a very high selectivity of this electrode for drug over most of the tested species. The results listed in Table 3 reveal that there were no significant interferences from any of the tested substances. Overall, the designed electrode is useful for the intended measurements.

**Table 3.** Potentiometric selectivity coefficients of some interfering ions

Interferent, J	$k_{midazolam_j}^{pot}$	Interferent,J	$k_{midazolam_j}^{pot}$
Ca <sup>+2</sup>	$2.5 \times 10^{-3}$	Fe <sup>+2</sup>	$5.5 \times 10^{-3}$
Pb <sup>+2</sup>	$4.1 \times 10^{-3}$	Al <sup>+3</sup>	$7.1 \times 10^{-3}$
Cd <sup>+2</sup>	$1.1 \times 10^{-3}$	Cr <sup>+3</sup>	$1.7 \times 10^{-3}$
Zn <sup>+2</sup>	$7.6 \times 10^{-3}$	sucrose	$4.39 \times 10^{-2}$
Ni <sup>+2</sup>	$1.01 \times 10^{-3}$	glucose	$4.78 \times 10^{-2}$
Cu <sup>+2</sup>	$8.5 \times 10^{-3}$	fructose	$4.78 \times 10^{-2}$
Ba <sup>+2</sup>	$1.5 \times 10^{-3}$	uric acid	$4.5 \times 10^{-2}$
Co <sup>+2</sup>	$6.1 \times 10^{-4}$		

### 3.8. Pharmaceutical preparations

Midazolam in pharmaceutical preparation (ampoule) were analyzed and their concentrations were determined using the proposed electrode. The results are listed in Table 4. Applying least-squares method for three determinations gave a 95% confidence level for the slopes and intercepts.

The regression line equation (taken versus found) for the proposed carbon paste electrode can be represented by:

$$Y=0.9865(\pm 0.0045) X - 0.0896(\pm 0.1325)$$

$$R=0.9999$$

Where X is the average reference assay and Y is the average of the proposed method. Correlation coefficient values of 0.9999 were obtained. The relative mean errors obtained ranged from 1.28 to 1.87% with % recovery from 98.16 to 98.71% (Table4).

These results indicate that the proposed electrode can be used to determine midazolam in pure samples or in pharmaceutical preparations with high accuracy, precision and high % recovery without pretreatment procedures of the samples to minimize interfering matrix effects.

**Table 4.** Determination of midazolam in pharmaceutical preparations using carbon paste electrode

Taken/ $\mu\text{g ml}^{-1}$	Midazolam carbon paste electrode MIDZCPE		Reference HPLC method	
	Found <sup>a</sup> / $\mu\text{g ml}^{-1}$	% Recovery	Found <sup>a</sup> / $\mu\text{g ml}^{-1}$	% Recovery
162.9	161.2	98.71	161.8	99.33
32.6	32.1	98.47	32.3	99.08
16.3	16.0	98.16	16.1	98.77

#### 4. CONCLUSION

In the present work, a novel modified carbon paste electrode was constructed for determination of midazolam. It offers a simple, accurate, selective, and specific tool for quantitative determination of midazolam content in its pharmaceutical preparation, midazolam ampoule. The sensor demonstrated advanced performances with a fast response time, a lower detection limit of  $6 \times 10^{-7}$  M and potential responses across the range of  $5 \times 10^{-3}$  -  $1 \times 10^{-5}$  M. The sensor enabled the midazolam determination in pharmaceutical formulations. This sensor respond based on ion-exchange mechanism. The best performance of carbon paste electrode was achieved by the ion-pair modifier 10%, Graphite powder 50% and plasticizer DOP 40%. A carbon paste electrode was designed to improve the analytical responses. And also a response of this developed method was compared with HPLC method.

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#### References

1. C. Page, C. Michael, M. Sutter, M. Walker, and B.B. Hoffman, *Integrated Pharmacology* (2nd ed.). C.V. Mosby Ltd., (2002), ISBN 978-0723432210.
2. K.T. Olkkola, and J. Ahonen, "Midazolam and other benzodiazepines". *Handbook of Experimental Pharmacology*, Springer, 182 (2008) 335.
3. A.C. Moffat, M.D. Osselton, and B. Widdop, *Clarke's Analysis of Drugs and Poisons*, 3<sup>th</sup> Edition, Pharmaceutical Press (2005).
4. M. Bodmer, B. Link, N. Grignaschi, O. Kummer, S. Ruegg, M. Haschke, S. Krahenbuhl, *Ther. Drug Monit.* 30 (2008) 120–124.
5. M. Pecking, F. Montestruc, P. Marquet, E. Wodey, M.C. Homery, P. Dostert, *Br. J.Clin. Pharmacol.* 54 (2002) 357–362.
6. J.G. Reves, R.J. Fragen, H.R. Vinik, D.J. Greenblatt, *Anesthesiology* 62 (1985) 310.
7. M. Gerecke, *Br. J. Clin. Pharmacol.* 16 (1983) 111.
8. H. Allonen, G. Ziegler, U. Klotz, *Clin. Pharmacol. Ther.* 30 (1981) 653.

9. J. Jurica, M. Dostalek, J. Konecny, Z. Glatz, E. Hadasova, J. Tomandl, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.* 852 (2007) 571–577.
10. P.G. ter Horst, N.A. Foudraïne, G. Cuyper, E.A. van Dijk, N.J. Oldenhof, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.* 791 (2003) 389–398.
11. V. Sautou, J. Chopineau, M.P. Terrisse, P. Bastide, *J. Chromatogr.* 571 (1991) 298–304.
12. V. Mastey, A.C. Panneton, F. Donati, F. Varin, *J. Chromatogr. B: Biomed. Appl.* 655 (1994) 305–310.
13. R. Lauber, M. Mosimann, M. Buhner, A.M. Zbinden, *J. Chromatogr. B: Biomed. Appl.* 654 (1994) 69–75.
14. F. Ma, C.E. Lau, *J. Chromatogr. B: Biomed. Appl.* 682 (1996) 109–113.
15. S.L. Eeckhoudt, J.P. Desager, Y. Horsmans, A.J. De Winne, R.K. Verbeeck, *J. Chromatogr. B: Biomed. Sci. Appl.* 710 (1998) 165–171.
16. M. Bodmer, B. Link, N. Grignaschi, O. Kummer, S. Ruegg, M. Haschke, S. Krahenbuhl, *Ther. Drug Monit.* 30 (2008) 120–124.
17. A. Petsalo, M. Turpeinen, O. Pelkonen, A. Tolonen, *J. Chromatogr. A* 1215 (2008) 107–115.
18. S.J. Marin, R. Coles, M. Merrell, G.A. McMillin, *J. Anal. Toxicol.* 32 (2008) 491–498.
19. B. Link, M. Haschke, M. Wenk, S. Krahenbuhl, *Rapid Commun. Mass Spectrom.* 21 (2007) 1531–1540.
20. F.E. Dussy, C. Hamberg, T.A. Briellmann, *Int. J. Legal Med.* 120 (2006) 323–330.
21. H. Kanazawa, A. Okada, E. Igarashi, M. Higaki, T. Miyabe, T. Sano, R. Nishimura, *J. Chromatogr. A* 1031 (2004) 213–218.
22. S.N. Muchohi, S.A. Ward, L. Preston, C.R. Newton, G. Edwards, G.O. Kokwaro, *J. Chromatogr. B: Anal. Technol. Biomed. Life Sci.* 821 (2005) 1–7.
23. J. Martens, P. Banditt, *J. Chromatogr. B: Biomed. Sci. Appl.* 692 (1997) 95–100.
24. G. Frison, L. Tedeschi, S. Maietti, S.D. Ferrara, *Rapid Commun. Mass Spectrom.* 15 (2001) 2497–2501.
25. M.J. Shao, K.D. Fallon, S.N. Khalil, E. Abouleish, *J. Chromatogr.* 345 (1985) 184.
26. Y. Umezawa, P. Buhlmann, K. Umezawa, K. Tohda, S. Amemiya, *Pure Appl. Chem.* 72(2000) 1851.
27. A.A. Salem, B.N. Barsoum, E.L. Izake, *Analytica Chimica Acta* 498 (2003) 79–91
28. M. R. Ganjali, B. Larijani, P. Norouzi, *Int. J. Electrochem. Sci.*, 7 (2012) 4822 – 4833
29. M.N. Abbas, G.A.E. Mostafa / *J. Pharm. Biomed. Anal.* 31 (2003) 819–826
30. P. R. Buck, and E. Lindner, *Pure Appl. Chem.* 66 (1994) 2527.
31. K. A. Singh and S. Mehtab, *Sens. Actuators B.* 123(2007) 429.
32. K. A. Singh, S. Mehtab, A. K. Jain, *Anal. Chim. Acta.* 575(2006)25.